2022 Update on Thrips and INSV

Daniel K. Hasegawa Research Entomologist USDA-ARS, Salinas CA

Pest Management Meeting 9/3/2022



Agricultural Research Service

2022 Update on Thrips and INSV

Thrips/INSV biology and monitoring

- Symptomology
- Thrips life cycle and transmission of INSV

Thrips/INSV epidemiology

- Discrepancies in distribution
- Thrips monitoring
- Temperature data

Thrips flower surveys: crops vs. non-crops

Immune priming

Receptor interference

INSV genome sequencing

INSV host range

- Top 10 hosts
- Top 10 hosts as thrips reproductive hosts
- Weed abatement (example)









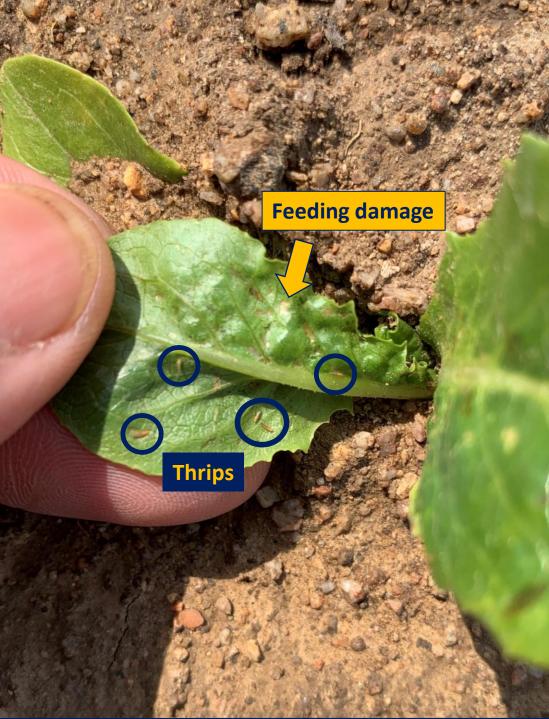






Western flower thrips: vector for INSV





Western flower thrips, Frankliniella occidentalis

Vector management challenges:

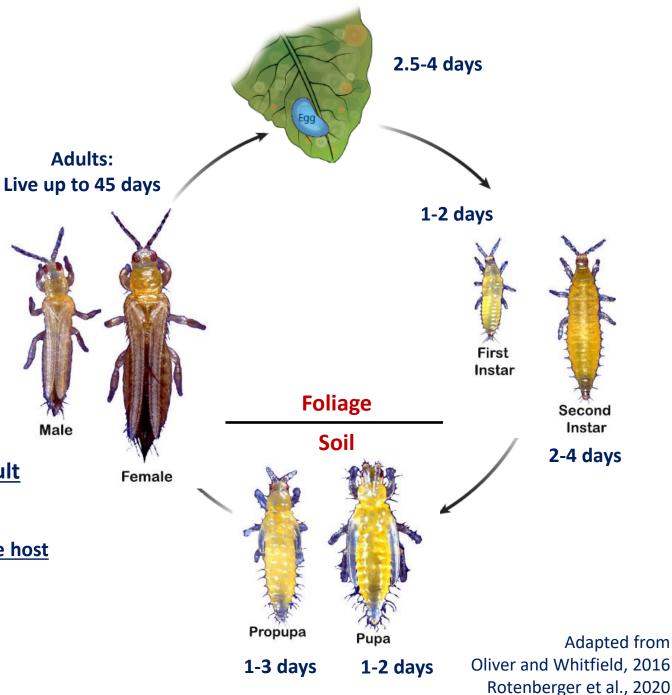
- Small (1-2 mm), cryptic, high fecundity
- Limited chemical options in CA lettuce
 - ~20% organic production in 2021
- Host range = 100s of plants

Virus Management challenges:

- Lack of genetic-based resistance to INSV in lettuce
- Host range = 100s of plants

Virus must be acquired as larvae to transmit as an adult

- Adults transmit the virus.
- Virus is not passed from adult to offspring.
- Plants that are infected with INSV must be a <u>reproductive host</u> for western flower thrips for virus acquisition to occur.





- INSV symptoms in lettuce: ~10-14 days
- Thrips larvae can acquire the virus before symptoms appear



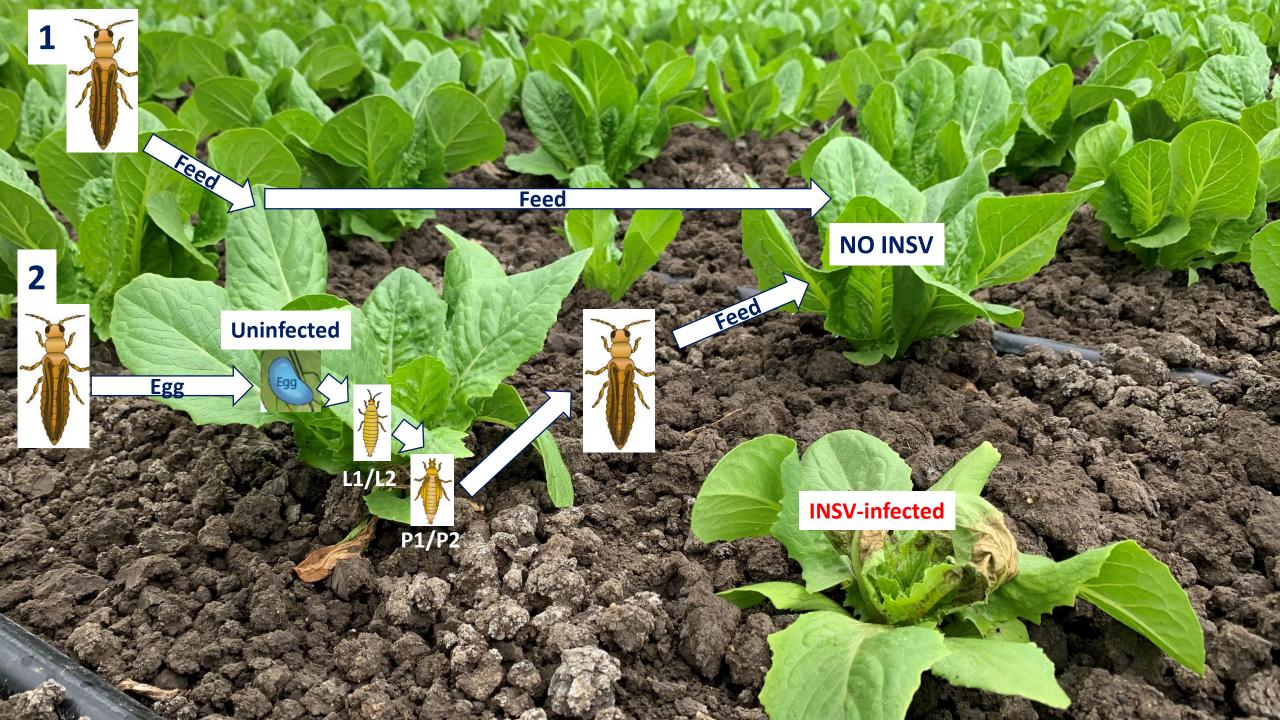


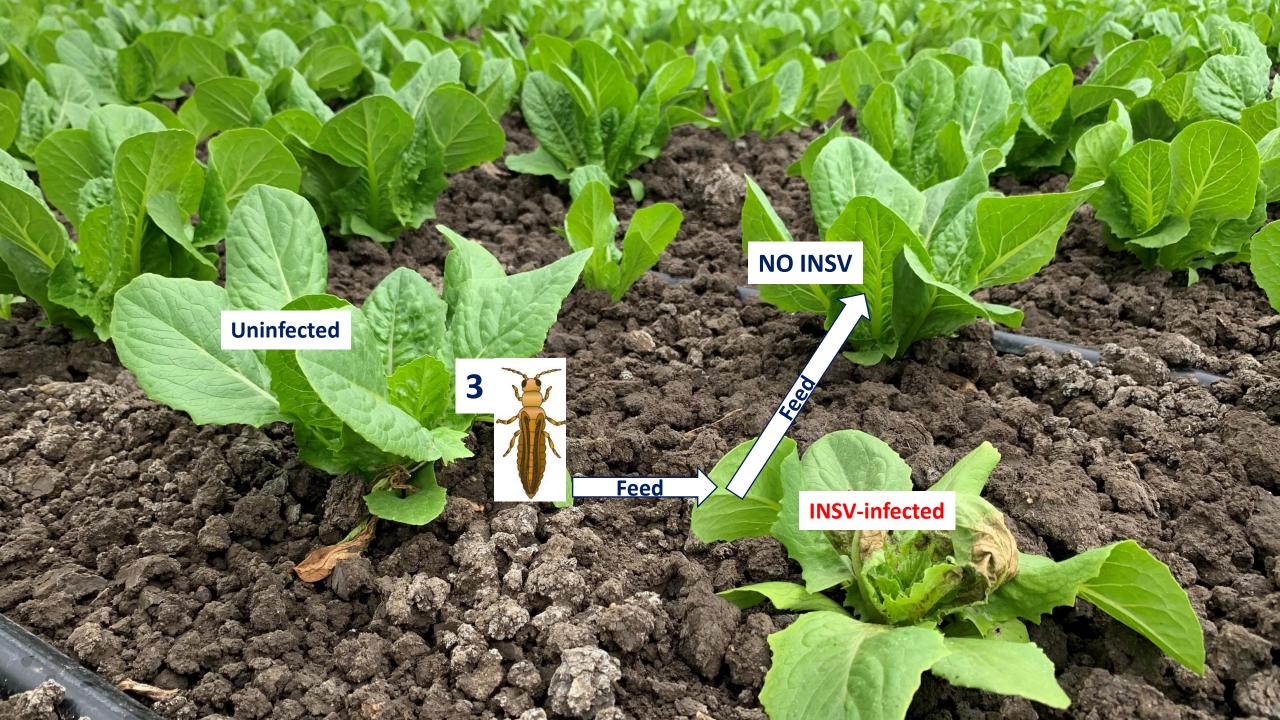
Thrips transmission of tospoviruses

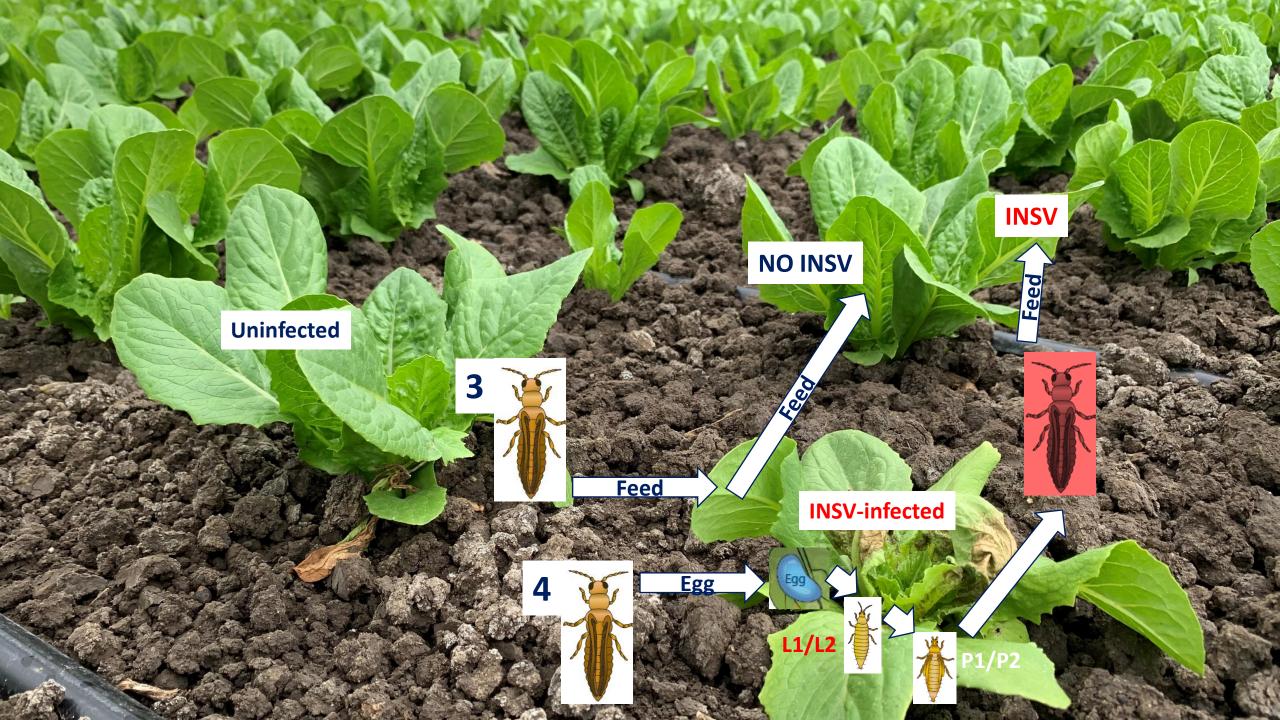
Uninfected

INSV-infected









Two types of infection

Secondary infection

NO

Primary infection

Outside field

2022 Update on Thrips and INSV

Thrips/INSV biology and monitoring

- Symptomology
- Thrips life cycle and transmission of INSV

Thrips/INSV epidemiology

- Discrepancies in distribution
- Thrips monitoring
- Temperature data

Thrips flower surveys: crops vs. non-crops

Immune priming

Receptor interference

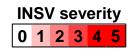
INSV genome sequencing

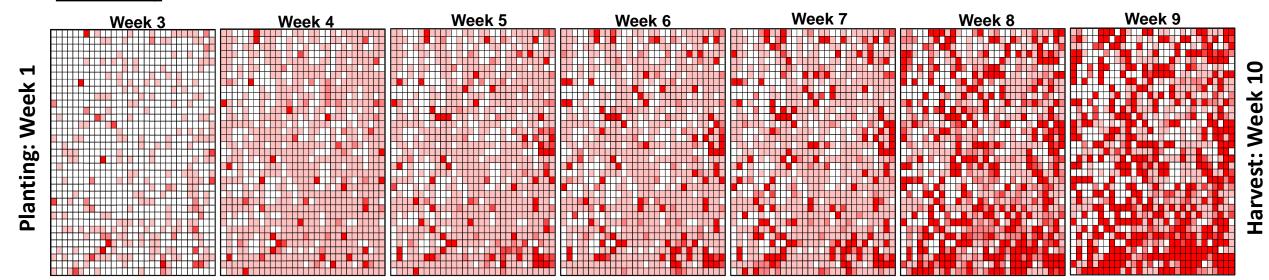
INSV host range

- Top 10 hosts
- Top 10 hosts as thrips reproductive hosts
- Weed abatement (example)

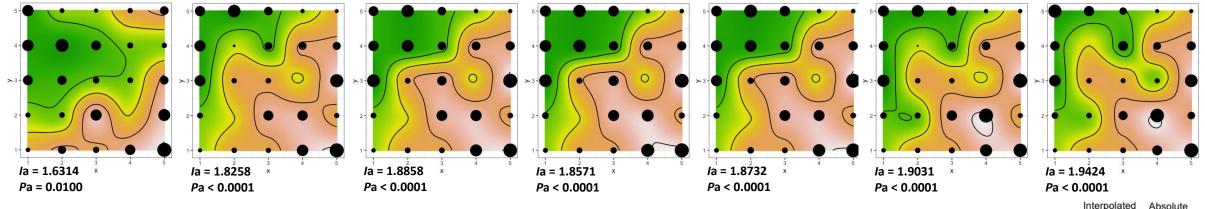
INSV Severity

INSV incidence strongly aggregates

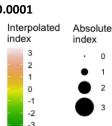




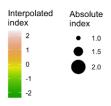
INSV Incidence



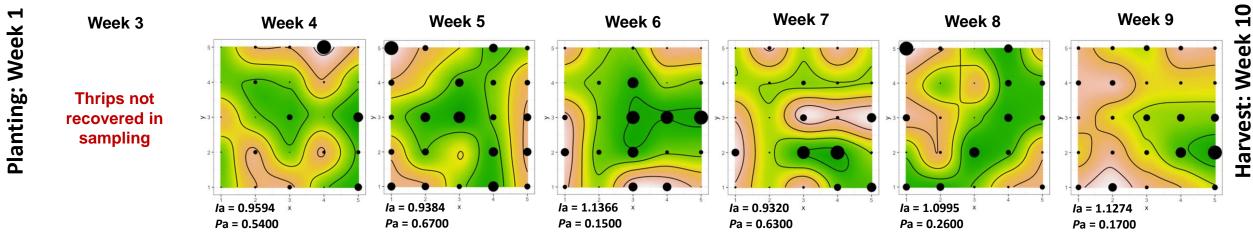




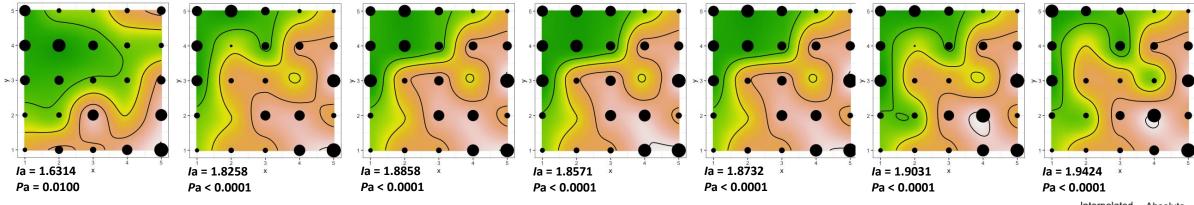
INSV incidence strongly aggregates

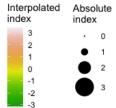


Thrips Abundance



INSV Incidence





Thrips distribution does not *always* equal INSV distribution

Hasegawa and Del-Pozo, 2022

Thrips abundance increases over time

 Interpolated index
 Absolute index

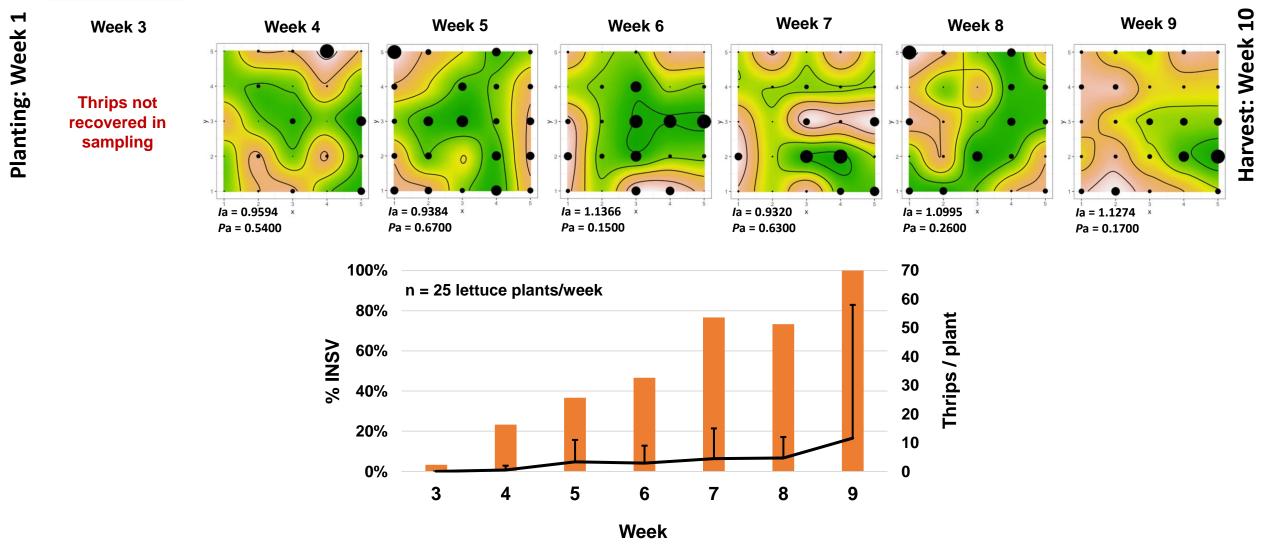
 2
 ● 1.0

 1
 ● 1.5

 0
 ≥.0

 -1
 -2

Thrips Abundance



Thrips abundance increases over time

 Interpolated index
 Absolute index

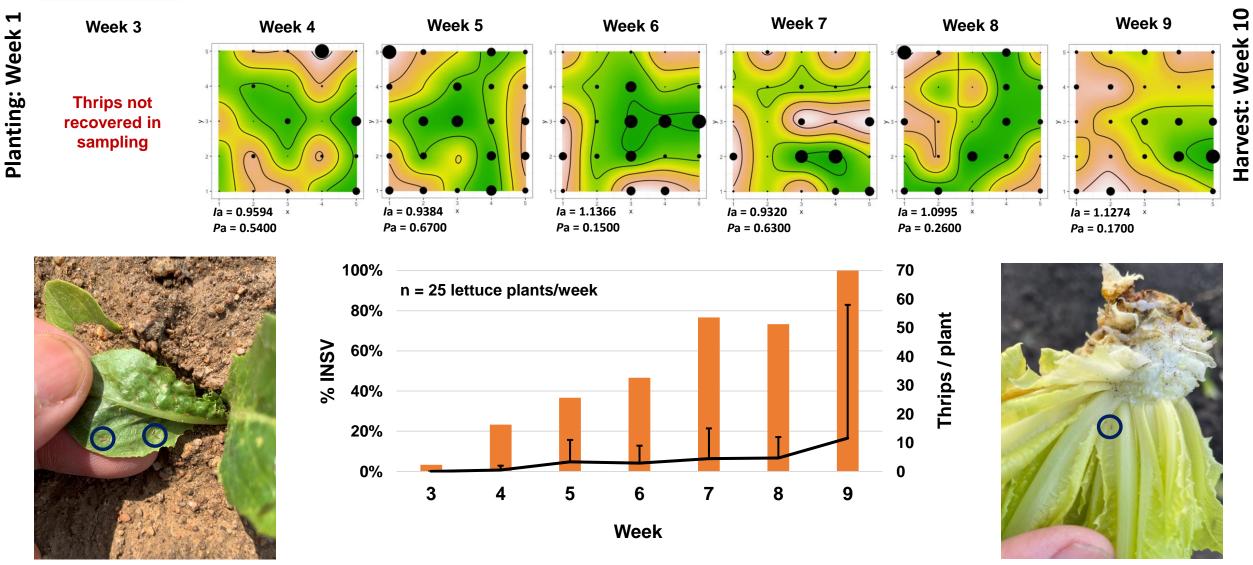
 2
 ● 1.0

 1
 ● 1.5

 0
 ● 2.0

 -1
 -2

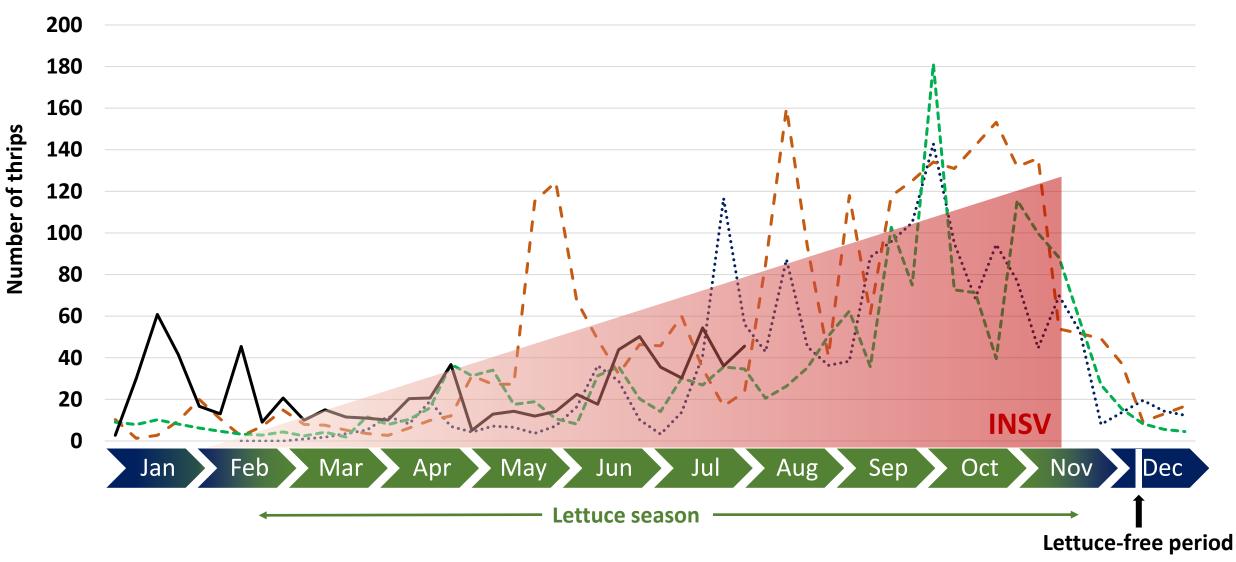
Thrips Abundance



Salinas Valley thrips monitoring network

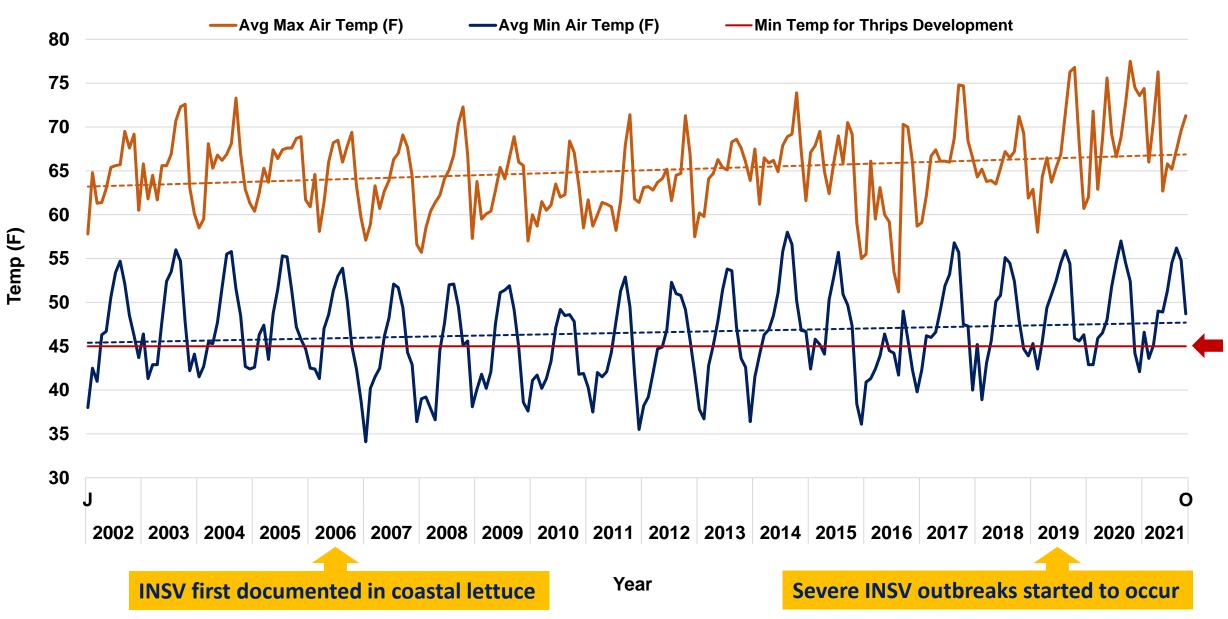
Thrips/sticky card/week (21 total, average)

······ 2019 - - 2020 --- 2021 --- 2022



Air temperature: 20 years

CIMIS Station 116: Salinas North



2022 Update on Thrips and INSV

Thrips/INSV biology and monitoring

- Symptomology
- Thrips life cycle and transmission of INSV

Thrips/INSV epidemiology

- Discrepancies in distribution
- Thrips monitoring
- Temperature data

Thrips flower surveys: crops vs. non-crops

Immune priming

Receptor interference

INSV genome sequencing

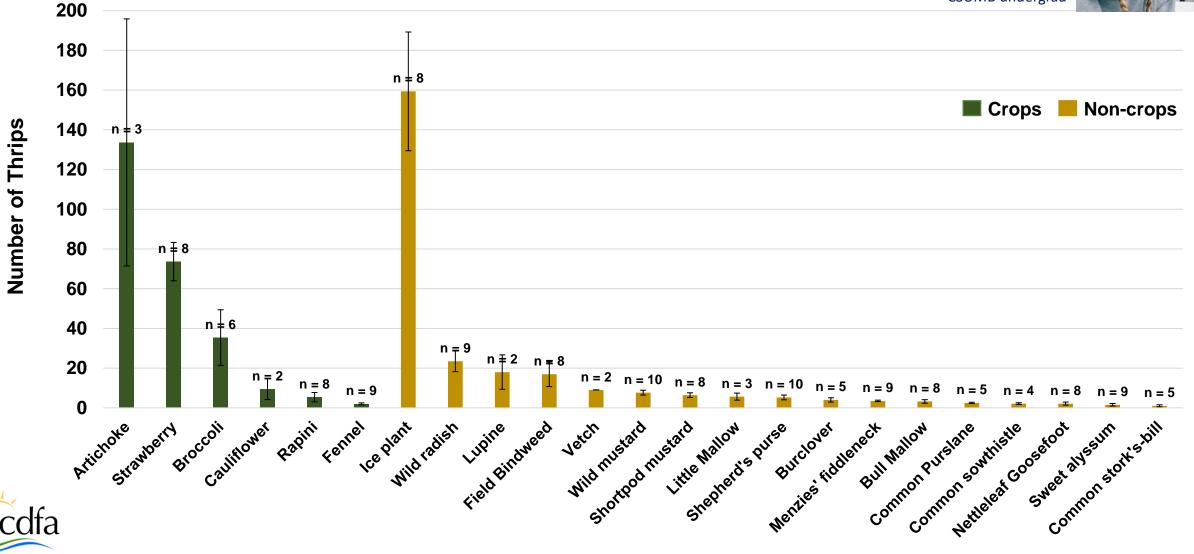
INSV host range

- Top 10 hosts
- Top 10 hosts as thrips reproductive hosts
- Weed abatement (example)

Thrips abundance in crops and non-crops

Average number of adult thrips/10 flowers

Kiara Gable USDA, Salinas CSUMB undergrad



CALIFORNIA DEPARTMENT OF FOOD & AGRICULTURE

2022 Update on Thrips and INSV

Thrips/INSV biology and monitoring

- Symptomology
- Thrips life cycle and transmission of INSV

Thrips/INSV epidemiology

- Discrepancies in distribution
- Thrips monitoring
- Temperature data

Thrips flower surveys: crops vs. non-crops

Immune priming

Receptor interference

INSV genome sequencing

INSV host range

- Top 10 hosts
- Top 10 hosts as thrips reproductive hosts
- Weed abatement (example)

Enhancing Virus Control in Lettuce and Melons by Optimizing Immunity Priming Approaches

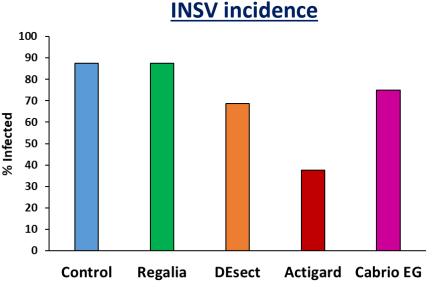
2.5 years (2021 – 2024); Year 1: Greenhouse trials

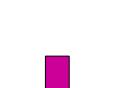
- 1. Actigard (AI = acibenzolar-S-methyl [ASM]).
- 2. Regalia (AI = extract of Giant Knotweed Reynoutria sachalinensis).

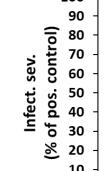
California Environmental Protection Agence

Department of

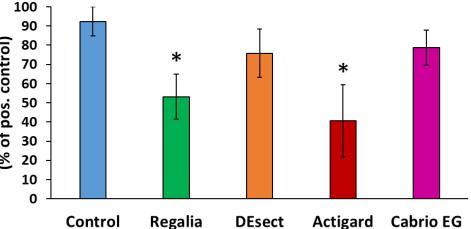
- **3.** Cabrio EG (AI = pyraclostrobin).
- 4. DEsect (AI = silicon dioxide).















Dr. Kerry Mauck Assistant Professor of Entomology, UC Riverside

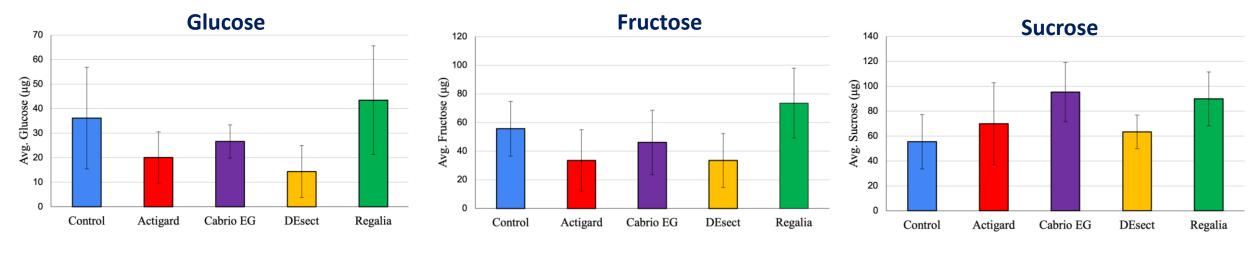
Enhancing Virus Control in Lettuce and Melons by Optimizing Immunity Priming Approaches

2.5 years (2021 – 2024); Year 1: Greenhouse trials

- 1. Actigard (AI = acibenzolar-S-methyl [ASM]).
- 2. Regalia (AI = extract of Giant Knotweed *Reynoutria sachalinensis*).
- **3.** Cabrio EG (AI = pyraclostrobin).
- 4. DEsect (AI = silicon dioxide).



Dr. Kerry Mauck Assistant Professor of Entomology, UC Riverside







California Environmental Protection Agency Department of Pesticide Regulation

Enhancing Virus Control in Lettuce and Melons by Optimizing Immunity Priming Approaches

2.5 years (2021 – 2024); Year 2: Field trial

- 1. Actigard (AI = acibenzolar-S-methyl [ASM]).
- 2. Regalia (AI = extract of Giant Knotweed *Reynoutria sachalinensis*).
- 3. Lannate SC (AI = Methomyl)
- 4. Radiant SC (AI = Spinetoram)
- 9 total treatments, 4 replicated blocks
- 3 applications, 2 weeks apart
- INSV incidence and severity, number of thrips recorded
- Data being collected



Dr. Kerry Mauck Assistant Professor of Entomology, UC Riverside





California Environmental Protection Agency Department of Pesticide Regulation

Receptor interference: A novel IPM technology for managing key insect pests of vegetables in California 2.5 years: (2022 – 2025)

- <u>Phase 1 (Discovery)</u>: Identification and expression of receptors that are specific to western flower thrips (WFT) and diamondback moth (DBM),
- <u>Phase 2 (Synthesis)</u>: Screen, design, and synthesize bioactive peptides that selectively bind to and disrupt WFT and DBM GPCRs, and
- <u>Phase 3 (Efficacy)</u>: Evaluate the efficacy of bioactive peptides on WFT and DBM survival.



PEST MANAGEMENT ...

New technology for environmentally safe pest control discovered

"Receptor interference" technology disrupts the vital processes needed for fire ants to survive



PUBLISHED ON AUGUST 29, 2021



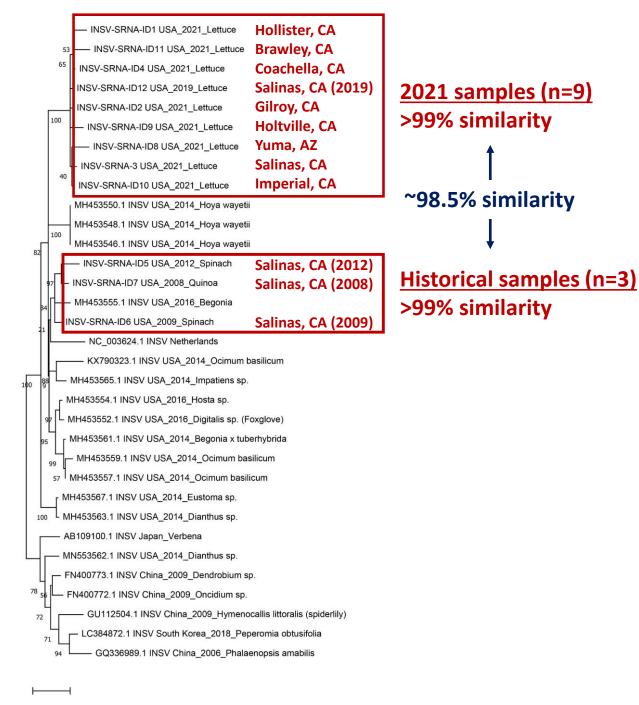
Dr. Manny Choi Research Entomologist, USDA Corvallis, OR



Laura Hladky USDA, Salinas Lab Tech



Juan Vargas USDA, Salinas CSUMB undergrad



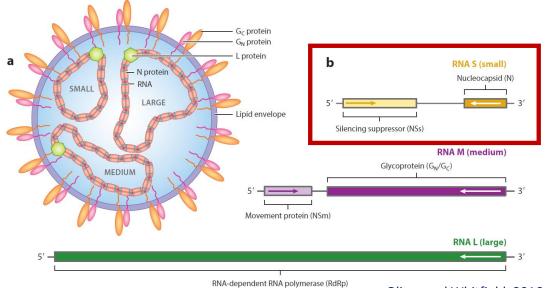
INSV genome sequencing

S RNA



Dr. Hanu Pappu **Distinguished Prof.** Plant Virology Washington State University

Dr. Ying Zhai Research Associate



Oliver and Whitfield, 2016

2022 Update on Thrips and INSV

Thrips/INSV biology and monitoring

- Symptomology
- Thrips life cycle and transmission of INSV

Thrips/INSV epidemiology

- Discrepancies in distribution
- Thrips monitoring
- Temperature data

Thrips flower surveys: crops vs. non-crops

Immune priming

Receptor interference

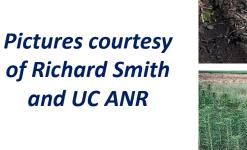
INSV genome sequencing

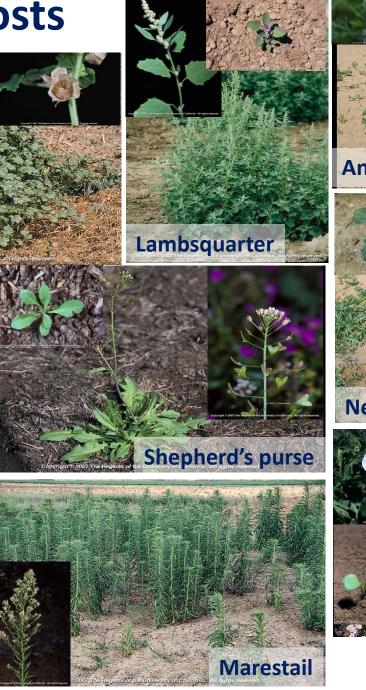
INSV host range

- Top 10 hosts
- Top 10 hosts as thrips reproductive hosts
- Weed abatement (example)

Top 10 hosts









Field Bindweed





Top 10 non-lettuce hosts for INSV on the Central Coast of CA

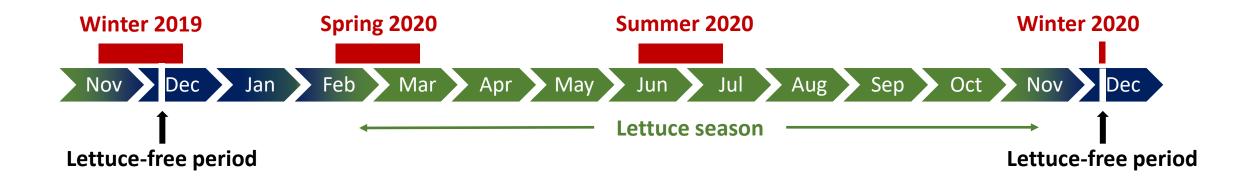
			Seasonal abundance				
Common name	Scientific name	Family	Category	Winter	Spring	Summer	Fall
Little Mallow	Malva parviflora	Malvaceae (Mallow Family)	Broadleaf	++	++	++	++
Annual Sowthistle	Sonchus oleraceus	Asteraceae (Sunflower Family)	Broadleaf	++	++	++	++
Nettleleaf goosefoot	Chenopodium murale	Chenopodiaceae (Goosefoot Family)	Broadleaf	+	++	++	++
Mare's Tail	Conyza canadensis	Asteraceae (Sunflower Family)	Broadleaf	+	++	++	++
Field Bindweed	Convolvulus arvensis	Convolvulaceae (Morning glory Family)	Broadleaf	0	++	++	++
Shepherds Purse	Capsella bursa-pastoris	Brassicaceae (Mustard Family)	Broadleaf	++	++	++	++
Common Purslane	Portulaca oleracea	Portulacaceae (Purslane Family)	Broadleaf	0	+	++	++
Hairy Fleabane	Conyza bonariensis	Asteraceae (Sunflower Family)	Broadleaf	+	++	++	++
Burning Nettle	Urtica urens	Urticaceae (Nettle Family)	Broadleaf	++	++	++	++
Common Lambsquarter	Chenopodium album	Chenopodiaceae (Goosefoot Family)	Broadleaf	0	++	++	++

Courtesy of Richard Smith

http://ipm.ucanr.edu/PMG/weeds_all.html

Field surveys to identify hosts for INSV: Salinas Valley

Sampling summary: >3,000 plant samples tested for INSV 73 species: majority weeds, native plants, vegetable crops



Primary detection of INSV: Serological: TAS-ELISA

Validation: Serological: Lateral flow rapid strip tests Genetic: RT-PCR





Laura Hladky USDA, Salinas Lab Tech

CALIFORNIA

RESEARCH PROGRAM

Ranking system for INSV hosts



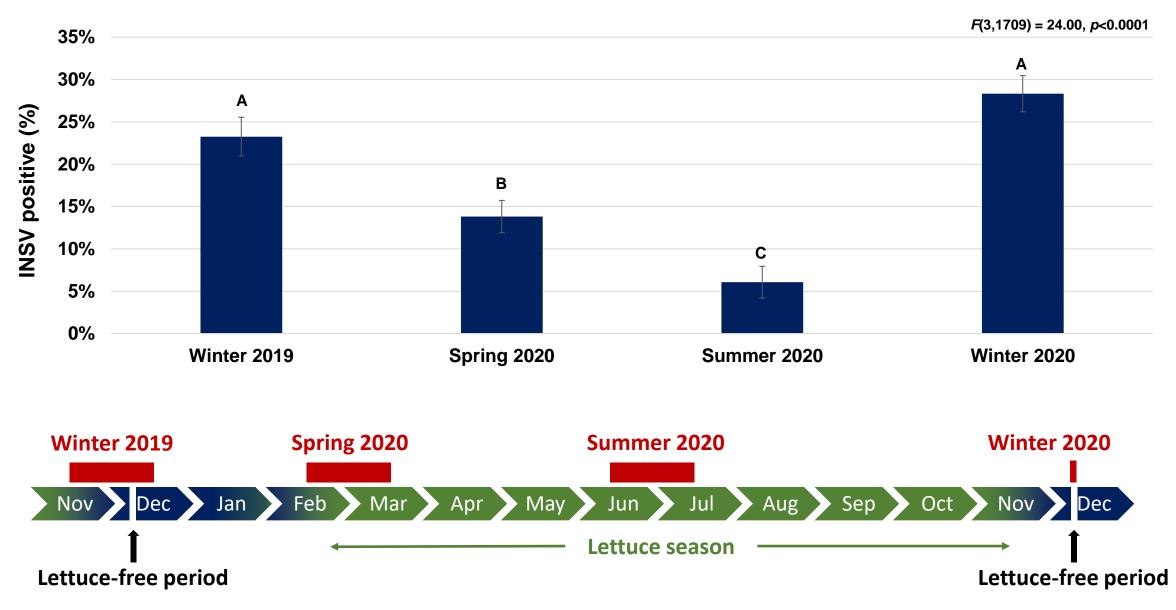
Host INSV Index = Avg ELISA_{positive} x (N_{positive}/N_{total})

Common name	Avg. ELISA Abs.	INSV positive	Total samples	% INSV	Host INSV	Validation:	Validation:	Validation:
	A48. EEISA A83.		iotal samples	/011131	Index	Immunostrip	RT-PCR	Seq. (partial)
Lettuce*	3.061	100	100	100.0	3.061	✓	✓	✓
Chickweed*	3.210	26	50	52.0	1.669	✓		
Hairy fleabane	1.730	26	93	28.0	0.484	✓		
Annual sowthistle	1.721	64	295	21.7	0.373	✓	✓	\checkmark
Common lambsquarter	1.211	15	54	27.8	0.336		✓	\checkmark
Purslane	1.724	13	75	17.3	0.299	✓	✓	\checkmark
Curly dock	1.508	9	46	19.6	0.295	✓	✓	✓
Field bindweed	1.137	26	118	22.0	0.251	✓	✓	✓
Shepherd's purse	2.557	11	116	9.5	0.242		✓	
Little mallow	1.066	101	544	18.6	0.198	✓	✓	✓
Mare's tail	1.720	15	157	9.6	0.164			
Bristly oxtongue	1.073	8	54	14.8	0.159			
Burning nettle	1.020	9	59	15.3	0.156		✓	✓
Redroot pigweed	1.592	2	30	6.7	0.106		✓	
Nettleleaf goosefoot	0.605	34	214	15.9	0.096	✓	✓	
Prickly lettuce	0.705	3	48	6.3	0.044	✓	✓	✓
Shortpod mustard	1.594	10	406	2.5	0.039	✓	✓	
Swine cress	0.644	1	30	3.3	0.021		*et	
Artichoke	0.261	1	30	3.3	0.009	4	the wat	Ne Ne J
Wild arugula	0.291	1	39	2.6	0.007		net nost at	o jet jet il
Bull mallow	0.946	1	131	0.8	0.007	iste pusion	in on critical and the	ed while the store of the state
Alkali mallow	0.249	0	31	0.0	0.000	it out out	inderetie using both the the	ed whether the structure whether the structu
Iceplant	0.249	0	91	0.0	0.000			
Field mustard	0.249	0	32	0.0	0.000			ē 👝
Plantain	0.249	0	31	0.0	0.000	-		Hasegawa

Grace Hardy USDA, Salinas CSUMB undergrad

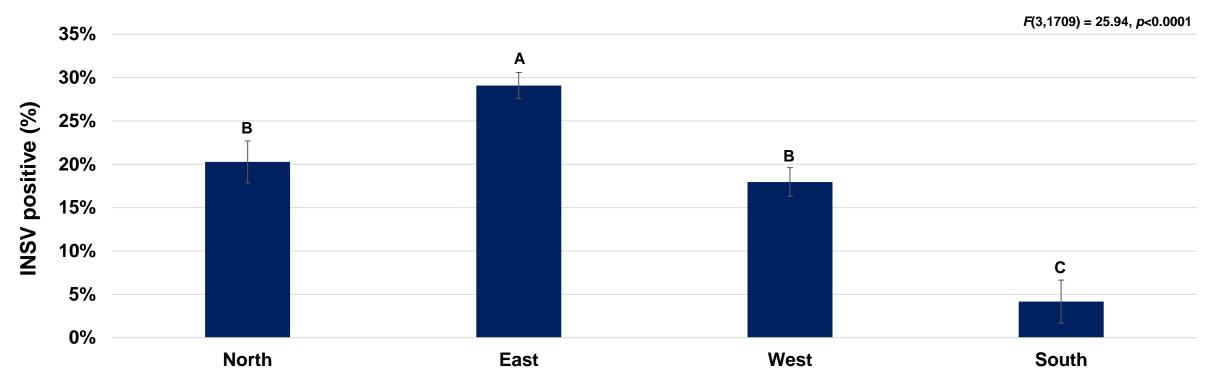
Hasegawa et al., in prep

Top 10 hosts: Season



Hasegawa et al., in prep

Top 10 hosts: Location







Hasegawa et al., in prep

Western flower thrips, Frankliniella occidentalis

Vector management challenges:

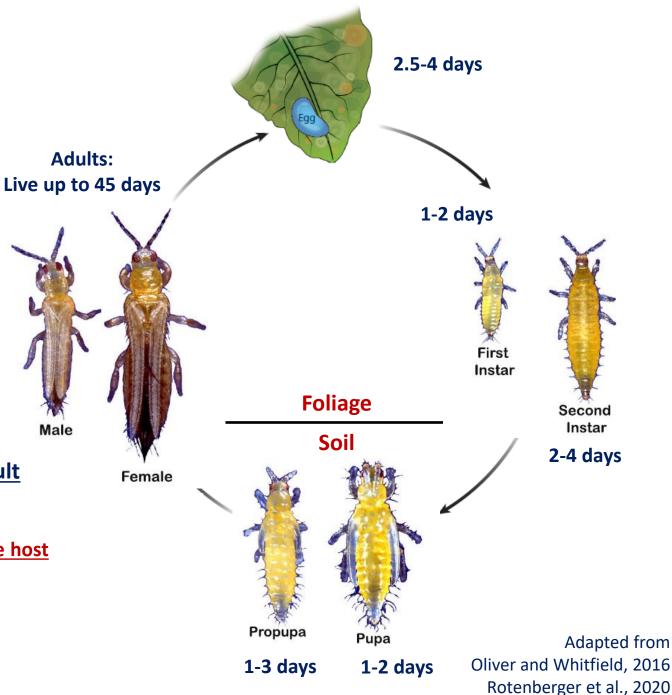
- Small (1-2 mm), cryptic, high fecundity
- Limited chemical options in CA lettuce
 - ~20% organic production in 2021
- Host range = 100s of plants

Virus Management challenges:

- Lack of genetic-based resistance to INSV in lettuce
- Host range = 100s of plants

Virus must be acquired as larvae to transmit as an adult

- Adults transmit the virus.
- Virus is not passed from adult to offspring.
- Plants that are infected with INSV must be a <u>reproductive host</u> for western flower thrips for virus acquisition to occur.

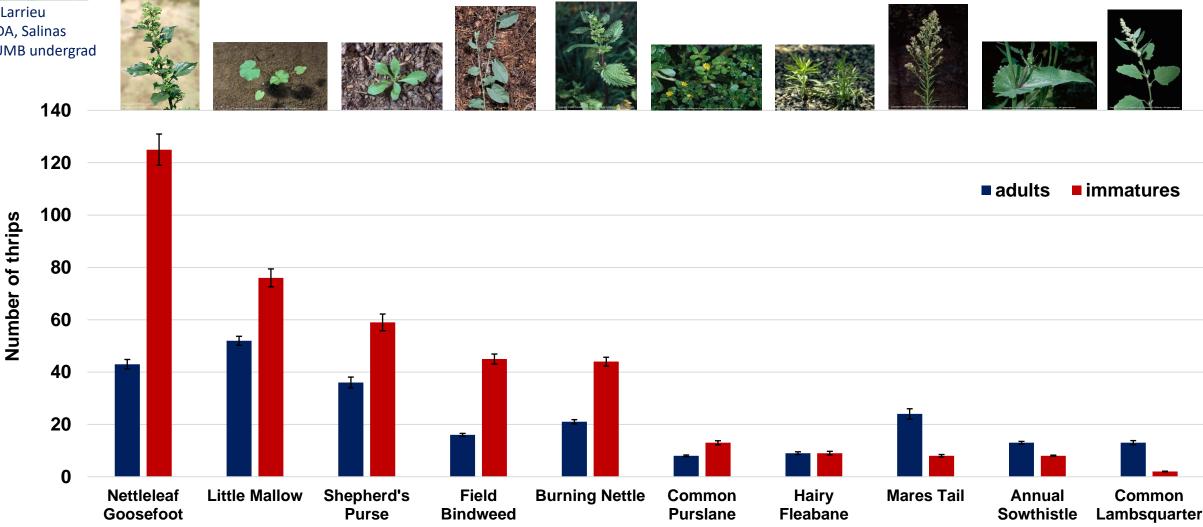




Kai Larrieu USDA, Salinas **CSUMB** undergrad

Thrips reproduction on top 10 hosts for INSV

Average number of thrips/plant; 10 plants per species



Focused weed management: Example

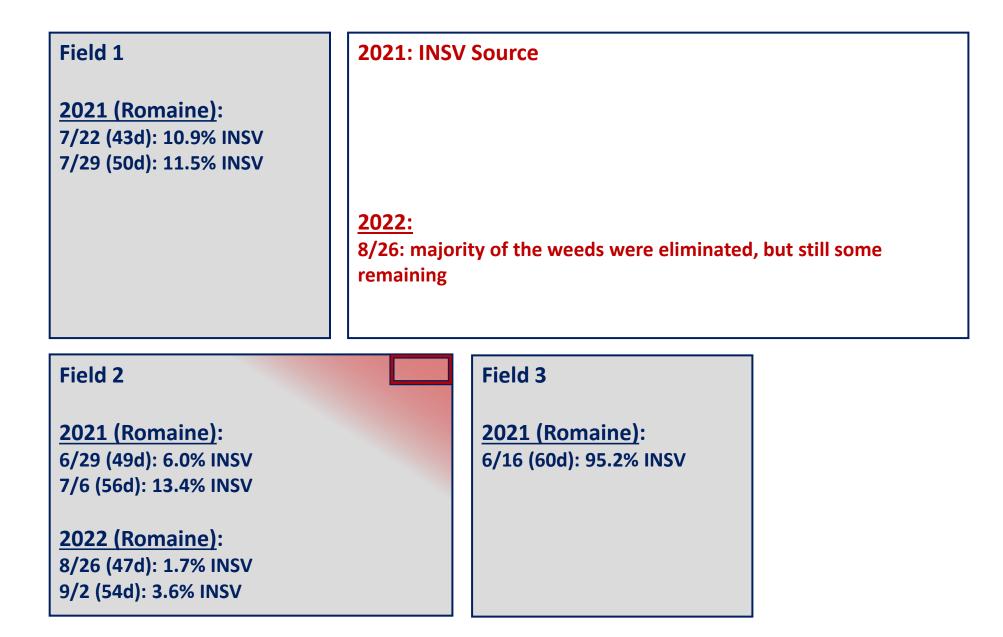
Field 1	2021: INSV Source		
<u>2021 (Romaine)</u> :	INSV positive / 9 Total		% INSV positive
	Little mallow	17/44	38%
	Annual sowthistle	23/37	62%
	Nettleleaf goosefoot	32/42	76%
	Common knotweed	8/36	22%

Field 3
<u>2021 (Romaine)</u> :

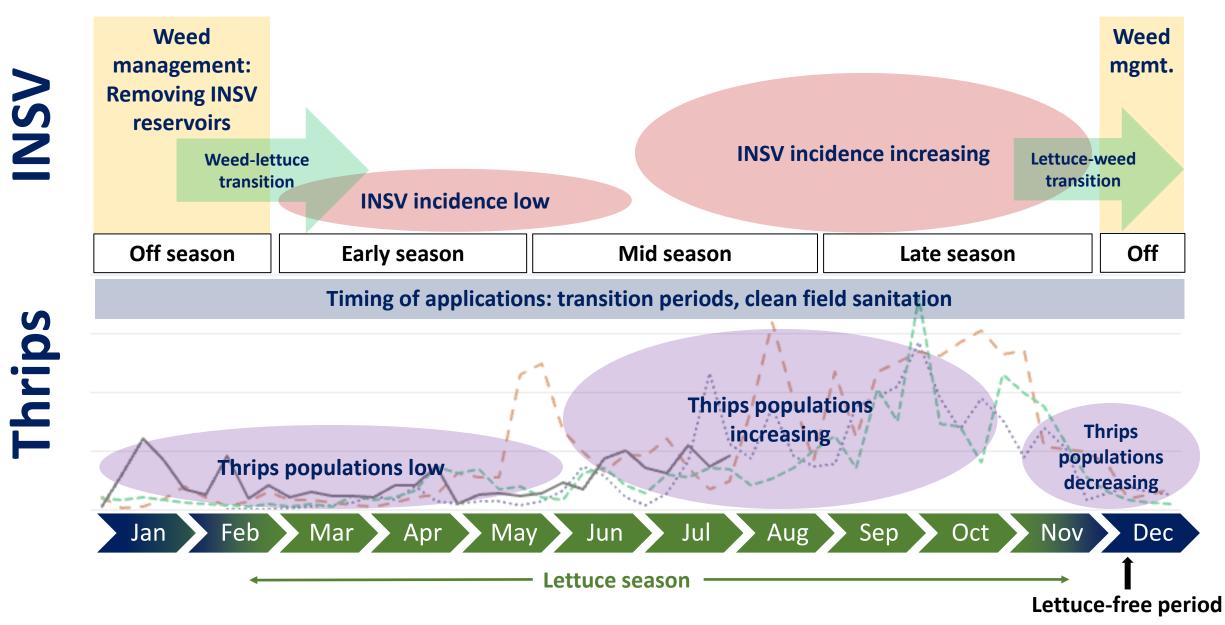
Focused weed management: Example

2021 (Romaine): 7/22 (43d): 10.9% INSV		INSV positive / Total	% INSV positive
7/29 (50d): 11.5% INSV	Little mallow	17/44	38%
	Annual sowthis	tle 23/37	62%
	Nettleleaf goose	efoot 32/42	76%
	Common knotw	veed 8/36	22%
Field 2	Fie	eld 3	
<u>2021 (Romaine)</u> :	20	21 (Romaine):	
6/29 (49d): 6.0% INSV	6/3	16 (60d): 95.2% INSV	
7/6 (56d): 13.4% INSV			

Focused weed management: Example



Thrips/INSV IPM model



Thank you

USDA-ARS Salinas, CA, Entomology Lab

Lab technician: Laura Hladky Students: Grace Hardy, Kiara Gable, Kai Larrieu, Jasmin Azad-Khan, Juan Vargas, Aaron Rocha

University of California:

Richard Smith (Vegetables and Weeds, Monterey County) Apurba Barman (Entomology, Imperial County)

Grower-Shipper Association of Central California Chris Valadez, GSA President Mary Zischke, INSV/Pythium Task Force leader

Growers and PCAs

USDA-ARS Salinas, Virology CA Bill Wintermantel, Plant Virologist Students: Aaron Rocha

Email: daniel.hasegawa@usda.gov Cell: 831-206-8177









CALIFORNIA DEPARTMENT OF FOOD & AGRICULTURE



